More agarics from xerophytic grasslands in central Spain

G. MORENO, F. ESTEVE-RAVENTÓS, C. ILLANA AND M. HEYKOOP

Department of Plant Biology (Botany), Alcalá de Henares University, Madrid, Spain

More agarics from xerophytic grasslands in central Spain. Mycological Research 94 (6): 781-788 (1990).

Agaricus goossensiae var. pseudolutosus var. nov., Conocybe cylindracea, Coprinus megaspermus, C. ovatus, C. vosoustii, Lepiota locquinii f. rioussetiae, Leucoagaricus macrorrhizus, Mycena chlorantha, M. olivaceomarginata and M. pseudopicta are reported from xerophytic grasslands in central Spain.

Key words: Agaricales, Agaricus, Conocybe, Coprinus, Lepiota, Leucoagaricus, Mycena, Systematics, Ecology, Xerophytic grasslands, Spain.

The fungi from xerophytic grasslands of the Iberian Peninsula have not been the subject of detailed study, probably due to the inconstancy of fruiting. This contribution follows an earlier account by Moreno & Esteve-Raventós (1988).

Agaricus goossenslae Heinem, var. pseudolutosus Moreno et al., var. nov. (Figs 1–7)

Etym.: Resembling A. lutosus according to the illustrations of Cappelli (1984)

A typo differt cystidiis nullis. Holotypus H.AH 11488.

Pileus 2·5-5 (-6·5) cm diam, hemispherical to convex, finally plano-convex, whitish to whitish-ochre, turning more or less deep yellow with age or when touched, smooth at first, then with concentric purplish squamules at the centre; margin incurved to straight, slightly floccose when young. Lamellae free, crowded, grey-pinkish to pink when young, purplegreyish when old, without a white edge. Stipe 2-4 × 0.4-0.7 cm, cylindrical, never bulbous, sometimes curved, not tapering at the base, whitish, discolouring more or less yellow when old or touched, specially at the base where it is slightly floccose; ring pendant, simple, more or less membranous, whitish, sometimes with brownish tints at the margin. Context whitish, turning slightly pinkish; odour fungoid, slightly of almonds; taste pleasant, sweetish. Schaeffer's reaction positive, even in dry specimens. Spores 6-7.5 × 4.5-5.5 (-6) µm, ovate, brown-purplish, smooth, thick-walled, without germ-pore. Basidia $25-30 \times 7-9 \mu m$, 4-spored, clavate. Lamella-edge fertile, with basidia and basidioles. Cystidia not observed. Pileipellis a cutis formed by elongated hyphae < 9 µm diam. Clampconnexions present.

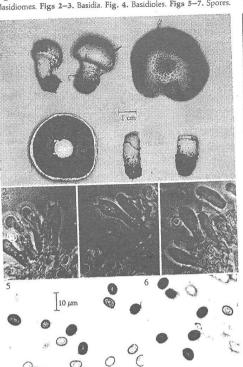
Specimens examined: Among Poaceae, in uncultivated, basic lands,

near Alcalá de Henares University, Madrid, 22 Oct. 1987, G. Moreno & F. Esteve-Raventós 10364; ibid., 10 Nov. 1986, V. Carnero, J. A. Esteban, A. Altés & G. Moreno 11488 (Holotype); ibid., 14 Nov. 1988, J. Alvarez, G. Moreno, A. Acha & M. Heykoop 11589; among Poaceae in acid soil, Arroyo del Barbaón, Natural Parc of Monfragüe, Cáceres, 3 Nov. 1987, G. Moreno, F. Esteve-Raventós & C. Illana 11582; among Poaceae in acid soil, Polideportivo de la Elipa, Madrid, 11 Nov. 1987, E. López 10487.

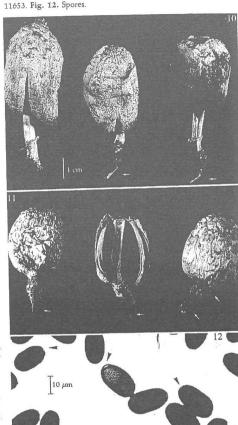
Agaricus goossensiae var. pseudolutosus is a member of Section Minores Fr., characterized by its small to medium size basidiome yellowing markedly, and with cap normally developing small purplish scales. The whole basidiome turns more or less yellow to orange-yellow with age or when touched, except the lamellae. Microscopically, the large spores and absence of true cheilocystidia are typical, and Schaeffer's reaction is positive which rules out the possibility of considering it among some yellowing taxa in the A. campester group. Agaricus lutosus Möller is very close to A. goossensiae var. pseudolutosus in its macroscopic characters, as can be observed in Capelli (1984), but this has smaller spores $(4-5 \times 3-3.5 \ \mu m)$, typical clavate cheilocystidia and tapering stipe. Owing to the absence of cheilocystidia, A. goossensiae var. pseudolutosus is closely related to A. comtulus Fr., but the latter has a completely white cap, never showing purplish colours, smaller spores (4.5–5.5 \times 3–3.5 $\,\mu m$), and hardly turns yellow with age or with friction. On the other hand, A. rusiophyllus Lasch, as conceived today, is a species with bulbous stipe, not turning so intensely yellow, as can be deduced in Capelli (1984) and the compilation by Bon (1985); furthermore, its spores are smaller (5.5 (-6) \times 4 μ m).

Agaricus goossensiae was proposed as a new species by Heineman (1956) from material collected in Zaire, but it is closest to the Spanish collections. It shares the same dimensions and colours of the basidiomes and spores, differing in the 782

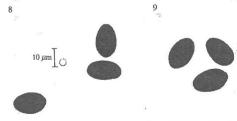
Basidiomes. Figs 2-3. Basidia. Fig. 4. Basidioles. Figs 5-7. Spores. 11653. Fig. 12. Spores.



Figs 1-7. Agaricus goossensiae var. pseudolutosus, 11488. Fig. 1. Figs 10-12. Caprinus avatus, 11654, 11653, Figs 10-11. Basidiomes;



Figs 8-9. Coprinus megaspermus, 11583, spores.



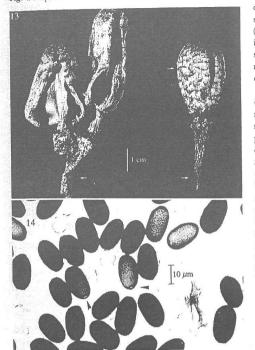
presence of cheilocystidia and the yellow pigmented lamellaedge. The African taxon was first described from dry material collected by Mme Goossens-Fontana in 1931. Heinemann (1956) points out that: 'la description est basée sur la planche et sur l'exsiccatum, l'état de ce dernier ne permet pas une étude

suffisante de l'hymenium'. On the other hand, one can deduce that this may be an abundant species in Zaire: 'comestible et de sayeur délicieuse, consommé par les populations Budjala et par les blancs'. The Spanish material could have been described as a new species, based on the microscopic characters, but it would be convenient to make further studies of this species in Africa to check the constancy of these differences.

Conocybe cylindracea Maire & Kühner ex Kühner & Watl., Notes Roy. Bot. Gard. Edinb. 40: 541 (1983).

This interesting Conocybe species is well characterized by its conico-ellipsoidal to cylindrical pileus; the stipe covered by hyaline hairs, devoid of lecythiform cystidia, ellipsoid spores with a conspicuous germ-pore and preference for mediterranean localities. It was originally described from Argelia (Kühner, 1935; Malençon & Bertault, 1970), and validated by Watling (1983); it was previously recorded in Spain by Moreno (1978).

Fig. 14. Spores.



Specimens examined: Among grass in an open and nitrogenous Ouercus suber forest, in acid soil, Crtra, de La Bazagona a Las Cansinas, Natural Parc of Monfragüe, Cáceres, 29 Oct. 1987, F. Esteve-Raventós & G. Moreno 10807; among Poaceae in grassland, in basic soil, Alcalá de Henares University, Madrid, 10 Nov. 1988, A. Altés, J. Esteban & G. Moreno 11581.

Coprinus megaspermus P. D. Orton, Notes Roy. Bot. Gard. (Figs 8-9) Edinb. 32: 141 (1978).

Our only collection agrees with the description of Orton & Watling (1979), and more recently, that of Uljé & Bas (1988). It shows the following main characters: pileus < 3 cm diam, stipe < 7 cm long, lamellae forming the typical 'C. plicatilis collarium', spores 15-19 × 8·5-11 µm, smooth, ellipsoid, very dark (nearly black), with an apical and central germ-pore, sometimes slightly lateral towards the abaxial face and cylindrical to lageniform marginal and facial cystidia.

Specimens examined: Among grass in an open Pinus sylvestris forest, in basic soil, Lagunillos, El Hosquillo, Cuenca, 7 May 1988, Gl. Moreno & G. Moreno 11583.

Coprinus ovatus (Schaeff.: Fr.) Fr., Epicr.: 242 (1838). (Figs 10-14)

Agaricus ovatus Schaeff., Fung. Bav. tab. 7 (1762). Agaricus ovatus Schaeff.: Fr., Syst. mycol. 1: 388 (1821). Coprinus comatus (Müll.: Fr.) S. F. Gray var. ? Malençon & Bertault, Fl. Champ. Sup. Maroc 1: 226-227 (1970).

Figs 13-14. Coprinus ovatus, 11652. Fig. 13. Basidiomes; 11653. Our material essentially agrees well with the description of Malencon & Bertault (1970); we have observed the following characters in our specimens; pileus reaching 5.5 × 4.5 cm; stipe always rooting markedly with a big mycelial cord (< 4 cm long); spores (13-) 13·5-19 × 8-11 μm, dark-brownish to purplish, with a central, apical germ-pore, sometimes slightly eccentric to its abaxial face; cheilocystidia globose, making the edge sterile; pleurocystidia not seen; clampconnexions present.

> Coprinus ovatus is different from the other species of Section Coprinus stirps comatus (Orton & Watling, 1979) by its rooting stipe and large spores with apical and central to slightly eccentric germ-pore. Coprinus comatus shares the scaly pileus and the lamellae turning pink before blackening but differs by its sporal range ($10-15 \times 6.5-8 \mu m$), never markedly rooting stipe, and more or less caespitose habit. Coprinus comatus forma sphaerocephalus J. Lange (Lange, 1939, pl. 156 fig. D), shows a thick brownish veil, small habit (cap originally almost globose, 5-6 cm) and nitrogenous habitat (collected in a park). His taxon might certainly be close to C. vosoustii Pilát; Lange's plate fits the macroscopic characters although he does not indicate the stellate disruption of the thick veil and spore morphology; we think that it should be considered a doubtful taxon. Coprinus sterquilinus (Fr.) Fr., is a fimicolous species with large spores, (16-) 17-22 × 10-13 (-13.5) µm (Orton & Watling, 1979), and shares similar habit and rooting stipe as C. ovatus. Coprinus oblectus (Bolt.) Fr., according to Fries' description, based on an illustration of Bolton, is a difficult taxon to interpret; it might be either C. vosoustii, C. ovatus or a form of C. comatus.

The only remarkable difference, pointed out in Fries' text, with our collected specimens would be that referred to the colour of the lamellae. C. ovatus sensu Fr. has white lamellae turning directly blackish at maturity without intermediate reddish or purplish colours. Schaeffer's plate no. 7 displays C. ovatus characters very well, closely resembling our material. According to our observations, C. ovatus is a species with mediterranean ecological preferences, found in xerophytic grasslands as well as in Quercus (Q. ilex, Q. rotundifolia and Q. suber) forests: this chorological data agrees with the remarks of Malencon & Bertault (1970).

Specimens examined: In humus of Quercus rotundifolia, in acid soil, El Pardo, Madrid, 11 Oct. 1986, F. Esteve-Raventós & G. Moreno 11652; in humus of Quercus suber, in acid soil, Exposición de Zafra, Badajoz, 23 April 1989, C. Sendín 11654; in xerophytic grasslands, in basic soil, Alcalá de Henares University, Madrid, 15 Nov. 1988, J. Alvarez, A. Acha, M. Heykoop & G. Moreno 11653, 11655.

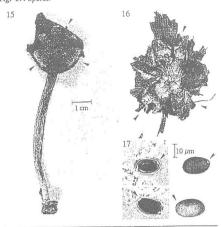
Coprinus vosoustii Pilát, Stud. Bot. Cech. 5: 207 (1942). (Figs 15-17)

Our collections macro- and microscopically agree very well with that of Donelli & Simonini (1989). Coprinus vosoustii is characterized by its ovate pileus smaller than in C. comatus; its typical brownish-ochre veil, thick, and persisting in a starshaped way after deliquescence; the bulbous, somewhat rooting stipe; and ellipsoid spores with eccentric germ-pore on the abaxial face. It seems to be a vernal species according to Donelli & Simonini (1989).

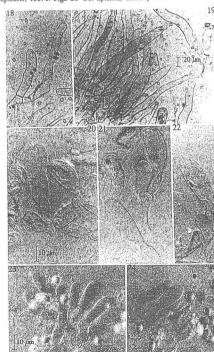
In Moreno (1976), this taxon was described as C. sterquilinus

784

Figs 15-17. Coprinus vosoustii, 1284. Figs 15-16. Basidiomes; 556. Fig. 17. Spores.



Figs 18-24. Lepiota locquinii f. rioussetiae, 10466. Figs 18-19. Epicutis; 10070. Figs 20-24. Epicutis, cheilocystidia.



var. dorsiporus var. nov., with the following characters: pileus 3-5 cm high, ovoid at first, then flat, with fibrous and furrowed margin, retaining a characteristic thick, brownish veil at the centre, which is not or hardly deliquescent. Stipe < 13 $\times\,0^{\circ}3-0^{\circ}5$ cm, white, cylindrical, hollow, fragile, somewhat bulbous and slightly rooting. Gills free, ascendant, deliquescent, white at first, turning pink, then blackish at maturity. Veil formed by filamentous hyphae. Spores 17- $23 \times 10-14$ µm, ellipsoid, smooth, brown-blackish, with a lateral, typically dorsal germ-pore at the abaxial face. Basidia 4-spored, clavate. Cheilocystidia globose to ellipsoid. Pleurocystidia not seen. Clamp-connexions present.

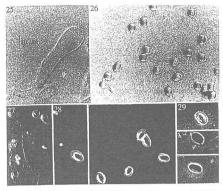
Specimens examined: In nitrogenous gardens, in acid soil, Facultad de Farmacia y Medicina, Madrid, 13 May 1976, K. Tabba 556; ibid., 19 May 1976, 8 May 1977 and 1 Oct. 1977, G. Moreno 1218, 1284,

Lepiota locquinii Bon forma rioussetiae Bon, Doc. Mycol. 61 - 46 (1985)

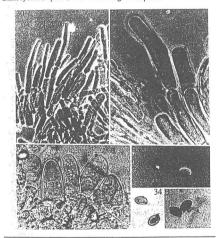
A small taxon, described by Bon (1985 b) and characterized by the lilac-pinkish tints on the stipe, and a trichodermium of variable hair-length. Our abundant collections, recorded for the first time in Spain and, for the second time, in Europe, show the following characteristics: pileus 0.6-2 cm diam, convex to plano-convex, uniformly argillaceous to brownishochre: epicutis breaking in very small squamules on a whitish ground, being strigose-floccose at the centre, which is concolorous or darker than the rest. Lamellae free, ventricose, whitish, with concolorous, floccose edge. Stipe 1-3 \times 0·1-0·3 cm, cylindrical, sometimes slightly enlarged at the base, not bulbous, whitish to argillaceous in its basal portion, with lilac to pinkish patent tints; ring white, located at the upper half of the stipe, very fugacious at maturity. Context whitish. Odour and taste absent or weak. Spores $6-9 \times 4.5-5.5$ µm, amygdaliform to broadly ellipsoid, more rarely subglobose, hyaline, smooth, dextrinoid, thin-walled, without germ-pore. Basidia 20-34 × 6-10 μm, 4-spored, more rarely 2-spored. Cheilocystidia very variable, clavate, cylindrical or fusiform, (15-) $18-25 \times 6-8$ µm. Subhymenium formed by catenulate, variable hyphae. Pileipellis a trichodermium of clavate to cylindrical hairs, 30–160 (–220) \times 10–15 (–20) μm , variable in length. Clamp-connexions present.

The pileal epicutis breaking at the centre in small strigose squamules, the ring present at first, the lilac to pinkish tints at the stipe and the epicutis formed by variable hairs are characteristic. The study of the holotype has revealed these macro- and microscopical similarities, though the spores of the type are smaller ($5.3-6 \times 3-4 \mu m$) yet morphologically similar. Spore variability has been frequently observed by us in the abundant material collected and examined; for instance, collections 10070 and 10071 show spores broadly ellipsoid to subglobose, 5–6 (–6·5) \times 4–5 $\,\mu m$, although all other characters are in agreement.

Specimens examined: Among Poaceae (mainly Dactylis glomerata ssp. hispanica), in basic soil, Alcalá de Henares University, Madrid, 26 Oct. 1987, F. Esteve-Raventós, G. Moreno, C. Illana & M. Heykoop 10469; in xerophytic grasslands under Salsola vermiculata, in basic soil, Finca de La Oruga, Alcalá de Henares, Madrid, 21 Nov. 1987, G. Moreno, F. Esteve-Raventós, C. Illana & M. Heykoop 10470, 77102002, Fig. 29, Spores.



Figs 30-35. Leucoagaricus macrorrhizus, 11622. Figs 30-34. Epicutis, cheilocystidia, spores; Bon 70307, Fig. 35, Spores.



10471, 10472 and 10473; same ecology and locality, 26 Oct. 1987, G. Moreno, F. Esteve-Raventós, C. Illana & M. Heykoop 10070, 10071; among Poaceae (mainly Dactylis glomerata spp. hispanica), in basic soil, Alcalá de Henares University, Madrid, 21 Nov. 1987, G. Moreno & F. Esteve-Raventós 10474.

Leucoagaricus macrorrhizus Locquin ex Horak, Beitr. Kryptogamenfl. Schweiz 13: 344 (1968). (Figs 30-35)

Leucocoprinus macrorrhizus Locquin, Bull. Mens. Soc. Linn. Lyon 12: 75 (1943).

Lepiota macrorrhiza (Locquin) Kühner & Romagn., Fl. Anal. Champ. Supér.: 406 (1953).

Though there are some discrepancies in the descriptions, our

Figs 25-29. Lepiota locquinii f. rioussetiae, 10070. Figs 25-27. material agrees well with most of them. Bon (1981, 1987) Finiculis, spores, basidium; 10466. Fig. 28. Spores; holotype Bon indicates and figures spores with a conspicuous germ-pore, but a study of material from his herbarium has revealed a small, hardly visible or absent germ-pore, very similar to those of our collection. It is an easily recognizable taxon, on the basis of its macroscopic characters: a more or less rooting stipe which turns brownish or red-brownish when touched or spontaneously when old, lamellae forming a 'collarium' near the stipe and a white-greyish to grey-ochre pileus with radially arranged fibres near the margin. However, we think that some characters can be very variable, such as the rooting stipe and the browning or reddening of the basidiomes. Wasser (1979) describes collections with rooting stipes, while Alessio (1988) indicates a hardly rooting stipe for the variety pinguipes. There are also some discrepancies about the presence of pleurocystidia, already indicated in Locquin's original description; they are present in our material, and Bon (1981) indicates the possibility of their presence, but nothing is said by Wasser (1979) or Donelli & Simonini (1984).

> Considered by Singer (1986) as the type species of this genus, L. macrorrhizus seem to show a certain preference for xerophytic areas: open and dry lands, more rarely in gardens. Wasser (1979) has found it commonly in the U.S.S.R. (Ukraine and Armenia) in dry steppes, and comments on the adaptation of the rooting stipe to dry lands. The distribution is limited to Europe and North Africa.

Alessio (1988) has recently considered L. pinguipes (Pearson) Bon to be a variety of L. macrorrhizus on the basis of their macroscopic characters, differing in the sandy maritime areas, and a scarcely rooting stipe.

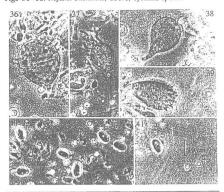
Specimens examined: In uncultivated lands barbechos, on basic soil, Guma, Burgos, 8 Oct. 1987, F. Esteve-Raventós 11622; in gardens, Charleville, France, Oct. 1970, Denoix 70307 (Herb. M. Bon).

Mycena chlorantha (Fr.: Fr.) Kummer, Führ. Pilzk.: 110 (Figs 36-41)

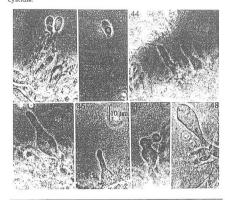
Our collection agrees well with the description of Maas Geesteranus (1984), except for the spores that are somewhat smaller. It represents a new record for the Iberian Peninsula and shows the following characters. Pileus -2 cm diam, hemisphaerical at first, then convex, green-olivaceous, hygrophanous and striate when moist. Lamellae ascendant, ventricose, adnate to emarginate near the stem, whitish with greenish tints at first, then grey with greenish-yellowish tints; edges yellow. Stipe -4.5×0.2 cm, cylindrical, smooth, slightly pruinose at the apex, greyish with greenish tints, the base strigose and covered by yellowish rhizoids. Taste none. Odour strongly of iodoform when old. Spores $8{\cdot}2{-}9\times4{\cdot}3{-}5{\cdot}2$ µm, ellipsoid, smooth, hyaline, amyloid. Basidia 4-spored, clavate. Cheilocystidia 25-45 × 10-23 µm, clavate, with apical, cylindrical excrescences of variable length (-8 µm), more or less distant. Pleurocystidia similar to cheilocystidia. Pileipellis with short excrescences. Context dextrinoid; clamp-connexions

Mycena chlorantha belongs in the Section Filipedes (Maas Geesteranus, 1980), and is easily recognized by the greenolivaceous colours of the basidiome, lamellae lacking pinkish tints, smell iodoform, and its ecology. Mycena flavescens Velen. is very close, but devoid of greenish colours and its smell is 786

Figs 36-41. Mycena chlorantha, 11478, cystidia, spores.



Figs 42-48. Mucena olivaceomarginata, 11675, basidium, spores, cvstidia.



raphanoid. Mycena arcangeliana (= M. oortiana auct.) is similar but shows pinkish tints at the lamellae at maturity and has a different ecology.

It is important to emphasize that our material was collected on buried stems of Poaceae in sandy soil, but not in coastal dunes, which seems to be the typical habitat of this species; nevertheless, Dr R. Courtecuisse kindly confirmed our determination as very typical. Courtecuisse & Guimbertau (1985) have recently described caespitose forms, indicating their frequency in the coastal dunes in northern France. Known in Europe from Denmark, France, Great Britain and The Netherlands; it has also been recorded recently in West Germany (Krieglsteiner, 1985). It always grows on Poaceae debris (Ammophila, Leymus, Carex, etc.) in sandy soils, normally in coastal dunes. A photograph has been recently published by Elborne (1989).

Specimens examined: On buried stems of Poaceae, in sandy soil, El Cabaco-La Alberca (Salamanca), 3 Dec. 1988, M. Ladero 11478.

Mycena olivaceomarginata (Massee) Massee, Br. Fung. Flora (Figs 42-48) 3: 116 (1893).

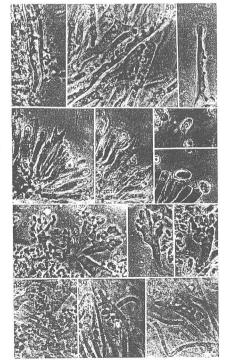
Mycena brunneomarginata Kühner, Encycl. Mycol. 10: 419

Mucena avenacea auct.

Highly variable in colour (Arnolds, 1982), this praticolous species of Section Rubromarginatae (Maas-Geesteranus, 1986), is not rare in the xerophytic grasslands of the Iberian Peninsula, However, it has only been recorded a few times before: Maire et al. (1933) and Maire (1937) from Catalonia. Losa-Ouintana & Freire (1978) from Galicia and Esteve-Raventós (1987) from Madrid, always as M. avenacea. Our specimens show a red to purplish lamella-edge and yellow to olivaceous stipe, which matches the strict sense of M. nlinaceomaroinata (Maas Geesteranus, 1986).

Specimens examined: Among Poaceae, in acid soil, Húmera, Madrid, 10 Nov. 1988. G. Moreno 11675.

Figs 49-60, Mucena pseudopicta, 11623, Figs 49-51. Epicutis. Figs 52-53. Bisporic basidia. Figs 54-55. Spores. Figs 56-57. Cheilocystidia. Figs 58-60. Caulocystidia.



Mycena pseudopicta (J. E. Lange) Kühner, Encycl. Mycol. 10: (Figs 49-60) 363 (1938).

Omphalina pseudopicta J. E. Lange, Dansk. bot. Ark. 6: 15

This is an interesting species with omphalinoid habit, which belongs to Section Cinerellae (Maas Geesteranus, 1986b), according to the subdivision of this genus made by Maas Geesteranus (1980). The species of this section are characterized by the omphalinoid habit, greyish to brownish colours, moist to viscid basidiomes, adnate to very decurrent gills, amyloid spores and cystidia with many coralloid excrescences. Mucena pseudopicta can be differentiated by its 2-spored basidia, absence of clamp-connexions, no particular smell, decurrent to triangular lamellae and very branched cystidia with irregular and large excrescences.

It seems to be distributed throughout Europe, although infrequent; it is known from Denmark, France, Greenland, Iceland, The Netherlands, Switzerland and West Germany; also from Spain, where it was previously recorded once by Mendaza & Díaz (1987) and north of Africa (Morocco) by Malencon & Bertault (1975). Complete descriptions and comments on this species can be found in Kühner (1938), Huiisman (1960) and Maas Geesteranus (1980, 1986b). Ecologically, M. pseudopicta is a graminicolous-heliophilous species, though it is not only restricted to grasslands; Courtecuisse (1984) has found it under shrubs and trees of various kinds.

Specimens examined: In xerophytic grasslands, among Poaceae, in basic soil, Alcalá de Henares University, Madrid, 10 Nov. 1988, J. Alvarez & C. Bartolomé 11623.

We wish to express our gratitude to H. Romagnesi, P. Heinemann (for his comments about Agaricus goossensiae var. pseudolutosus). M. Bon (for his revision of Lepiota locauinii forma rioussetiae and for material from his herbarium) and R. Courtecuisse (for his revision of Mycena chlorantha). We also thank the Ministerio de Educación y Ciencia for the concession of the Research Project No. PA 86-0063 to carry on with this work.

REFERENCES

ALESSIO, C. L. (1988). Leucoagaricus macrorrhizus Locq. ex Horak var. pinguipes (Pearson) n. comb. Micologia Italiana 2, 7-11.

ARNOLDS, E. (1982). Ecology and coenology of macrofungi in grasslands and moist heathlands in Drenthe, the Netherlands, Part 2 and 3. Bibliotheca Mycologica 90, 1-501.

BON, M. (1981). Clé monographique des 'Lepiotes' d'Europe (Agaricaceae, Tribus Lepioteae et Leucocoprineae). Documents Mycologiques 43, 1-77.

BON, M. (1985 a). Clé monographique du genre Agaricus L.: Fr. Documents Mycologiques 60, 1-37.

BON, M. (1985b). Novitates. Documents Mycologiques 61, 46.

BON, M. (1987). The Mushrooms and Toadstools of Britain and Northwestern Europe, London: Hodder & Stoughton.

CAPELLI, A. (1984). Fungi Europei. Agaricus L.: Fr. (Psalliota Fr.). Biella Giovanna, Saronno.

COURTECUISSE, R. (1984). Transect mycologique dunaire sur la cote d'Opale (France), (lere Partie: Les groupements héliophiles et arbustifs de la xerosère). Documents Mycologiques 57-58, 1-115.

COURTECUISSE R & GUIMBERTEAU, I. (1985), Mycena chlorantha fo. caespitosa Courtecuisse & Guimberteau fo. nov. Documents Mycologiques 61, 51-54.

DONELLI, G. & SIMONINI, G. (1984). Un fungo raro e amanti dei periodi siccitosi: Leucoggaricus macrorrhizus (Locg. ex Horak) Sing. Bolletino del Gruppo micologico G. Bresadola, Trento 27, 183-185.

DONELLI, G. & SIMONINI, G. (1989). Coprinus vosoustii Pilát e Coprinus phlyctidosporus Romagn. var. monobisporus Donelli et Simonini, XIII Mostra Reggiana del Fungo. 1, 2 et 3 Ottobre 1988. Assoc, Micol, Bresadola, Gruppo "R. Franchi"; 11-21. Reggio Emilia. ELBORNE, S. A. (1989). Danske Klitsvampe. Svampe 19, 1-11.

ESTEVE-RAVENTÓS, F. (1987). Contribución al conocimiento taxonómico, ecológico y corológico del orden Agaricales s.1. (Basidiomycotina) en el Sistema Central (Madrid y Segovia). Tesis Doctoral, Universidad de Alcalá de Henares (Madrid).

HEINEMANN, P. (1956). Flore Iconographique des Champignons du Congo. 5º fascicule: Agaricus I. Jardin Botanique de l'état, Bruxelles. HUIJSMAN, H. S. C. (1960), Mycena miserior Huijsm., spec. nov. et Mucena pseudo-picta (J. Lange) Kühn. Persoonia 1, 331-334.

KRIEGLSTEINER, G. J. (1985). Uber neue, seltene, kritische Makromyzeten in der Bundesrepublik Deutschland (Mitteleuropa). VI. Zeitschrift für Mykologie 51, 85-130.

KÜHNER, R. (1935). Le genre Galera. Encyclopédie Mycologique 7,

KÜHNER, R. (1938). Le genre Mycena. Encyclopédie Mycologique 10, 1-710

LANGE, J. E. (1939). Flora Agaricina Danica 4. Copenhagen.

LOSA-QUINTANA, J. M. & FREIRE, L. (1978). Macromicetes en Galicia: otoño 1975-invierno 1976. Braña, Boletín de la Sociedad Gallega de Historia Natural 1, 50-74.

MAAS GEESTERANUS, R. A. (1980). Studies in Mycenas-15. A tentative subdivision of the genus Mycena in the Northern Hemisphere. Persoonia 11, 93-120.

MAAS GEESTERANUS, R. A. (1984), Conspectus of the Mycenas of the Northern Hemisphere-3, Section Filipedes, Proceedings of the Koninklijke Nederlandse Akademie van Wetenschappen C 87,

MAAS GEESTERANUS, R. A. (1986a), Conspectus of the Mycenas of the Northern Hemisphere-8. Sections Intermediae, Rubromarginatae. Proceedings of the Koninklijke Nederlandse Akademie van Wetenschappen C 89, 279-310.

MAAS GEESTERANUS, R. A. (1986 b). Conspectus of the Mycenas of the Northern Hemisphere-7. Section Cinerellae. Proceedings of the Koninklijke Nederlandse Akademie van Wetenschappen C 89. 183-201

MAIRE, R. (1937). Fungi Catalaunici. Contribution a l'étude de la Flore Mycologique de la Catalogne. Publicacions de l'Institut Botanic, Barcelona 3, 1-128.

MAIRE, R. CODINA, J. & FONT QUER, P. (1933). Fungi Catalaunici. Contributions a l'étude de la Flore Mycologique de la Catalogne. Treballs del Museo de Cienciès Naturals de Barcelona, Ser. Botánica 15,

MALENCON. G. & BERTAULT, R. (1970-1975). Flore des Champignons Supérieurs du Maroc. Tomes 1 et 2. Institut Scientifique Chérifien et Faculté des Sciences. Rabat.

MENDAZA, R. & DIAZ. E. (1987). Las setas. Guía fotográfica y descriptiva, Iberduero, Vizcava,

MORENO, G. (1976). Estudio sistemático, ecológico y corológico del orden Agaricales (Basidiomycetes) en España. Tesis Doctoral, Universidad Complutense de Madrid.

MORENO, G. (1978). Algunas especies del género Conocybe Fayod que fructifican en los jardines y parques de la provincia de Madrid. Acta Botánica Malacitana 4, 5-10.

MORENO, G. & ESTEVE-RAVENTÓS, F. (1988). Agarics from

xerophytic grasslands in Central Spain. Transactions of the British Mycological Society 90, 407–413.

ORTON, P. D. & WATLING, R. (1979). British Fungus Flora. Agarics and Boleti 2. Coprinaceae: Coprinus. Edinburgh: Royal Botanic Garden.

SINGER, R. (1986). The Agaricales in Modern Taxonomy, 4th edn, J. Cramer. Vaduz.

ULJÉ, C. B. & BAS, C. (1988). Studies in Coprinus-1. Subsections

(Received for publication 25 July 1989)

Auricomi and Glabri of Coprinus section Pseudocoprinus. Persoonia

WASSER, S. P. (1979). Fungorum Rariorum Icones Coloratae. Pars X. J. Cramer. Vaduz.

WATLING, R. (1983). Observations on the Bolbitiaceae: 22. Further validations. Notes from the Royal Botanic Garden of Edinburgh 40, 537–558.

MYCOLOGICAL RESEARCH - advice to authors

The journal publishes Papers, Short Communications, and Reviews

SUBMISSION OF MANUSCRIPTS

Send

- (1) Two copies of the typescript only (no more) which must be double-spaced throughout.
- (2) Original line drawings (unlettered) plus two labelled copies, no larger than A4 size.
- (3) One unmounted print of each photograph plus two roughly mounted lettered prints, no larger than A4 size.
- (4) One copy of each paper cited as 'in press'.
- (5) Name and address of the person with whom the editors should correspond.

To: Dr David Moore

Department of Cell and Structural Biology Stopford Building The University Manchester M13 9PT U.K.

GENERAL

Mycological Research accepts papers in all fields of mycology, including Biotechnology and Industrial applications of fungi, Ecology, Genetics, Molecular Biology, Evolution, Plant and Animal Pathogens, Mutualistic Interactions, Physiology and Systematics. Review articles will be especially welcome. Manuscripts from nonmembers of the British Mycological Society will be considered for publication.

In the case of multiple authorship, it is assumed that all authors have consented to submission of the manuscript. Final acceptance is conditional on all authors signing the Copyright Assignment Form, which is issued with notification of acceptance.

By adhering to the following instructions precisely, authors will save editors and printers much unnecessary work and avoid delay in publication of their papers.

LAYOUT AND HEADINGS

Contributors must be clear and concise. There is no specific limit to length of papers.

The first page should give:

- (1) Title of the paper, which must be brief but accurate. Avoid 'generic' titles such as 'Studies of the physiology of yeast. XXIV.'
- (2) Names of authors and their affiliations.

- (3) A running title (not more than 50 characters and spaces).
- (4) A short summary.
- (5) A list of up to 7 suggested keywords.

N.B. The text should commence on the second page of the typescript.

Experimental papers should be set out with headings set left in bold face capitals – Materials and Methods, Results, Discussion. Primary subheadings should be set left in bold face italic and secondary subheadings, if required, in bold face italic at the left side of the page and indented. Editors recognize that not all work is best presented thus, so there is flexibility in layout. Footnotes should be avoided wherever possible. Pages should be numbered sequentially.

Line, half-tone and colour illustrations can only be accepted when necessary for the proper presentation of material.

The format for Short Communications is similar except that main and subheadings are generally discouraged.

Full instructions to authors and a specimen paper are given in Mycol. Res. 94(1): 136–141 (1990). Copies are available from the Executive Editor, Dr David Moore (Dept of Cell and Structural Biology, Stopford Building, The University, Manchester M13 9PT, U.K.) or the Production Editor, Dr B. C. Sutton (CAB International Mycological Institute, Ferry Lane, Kew, Richmond, Surrey TW9 3AF, U.K.).

PROCEDURES FOR PROOFS AND REPRINTS

Following acceptance of a manuscript for publication the author will receive a proof from the Printing Division of Cambridge University Press. Authors are asked to comply with the instructions accompanying the proofs explicitly. After correction the proof must be returned with the artwork (the originals and the material prepared by the printer) to Dr David Moore. The reason for this is that should additional correction be required to illustrations at second proof stage then the material will be immediately available. This will avoid the possibility of delay in publication.

Orders for reprints are handled by the printer (Cambridge University Press, Journals Publishing Dept, The Edinburgh Building, Shaftesbury Road, Cambridge, CB2 2RU, U.K.) not either the Executive or the Production Editors. Enquiries are to be directed accordingly.

The Editorial team of Mycological Research will appreciate the assistance of authors in following this advice,